**Simultaneous Purification of Genomic DNA and Total RNA from Animal Tissues**

Important points before starting:

* Do not allow tissue to thaw during weighing or handling prior to homogenization in Buffer RLT Plus/ß-Mercaptoethanol/Reagent DX mixture.
* Homogenized tissue lysates from step 4 can be stored at -70˚C for months.
* Buffer RLT Plus may form a precipitate upon storage. If necessary, redissolve by warming, and, then, place at room temperature.
* Preparation of Buffer RLT Plus/ß-Mercaptoethanol/Reagent DX mixure:
  + 10µL of 2-Mercaptoethanol is added to 1mL of Buffer RLT Plus prior to its use.
  + Reagent DX is added to Buffer RLT Plus at a final concentration of 0.05% (v/v)
* Buffer RPE, Buffer AW1, and Buffer AW2 are, each, supplied as a concentrate. Before using for the first time, add the appropriate volume of 100% ethanol, as indicated on the bottle, to obtain a working solution.
* Buffer RLT Plus, Buffer RW1, and Buffer AW1 contain a guanidine salt and are, therefore, not compatible with disinfecting reagents containing bleach.
* RNA purification requires proper aseptic technique for high-quality extracts.
* Place Buffer EB in lab coat pocket prior to beginning procedure to ensure gently warming of solution prior to DNA elution.
* Ensure Buffer RLT Plus/ß-Mercaptoethanol/Reagent DX mixture does not freeze in dry ice-cooled lysing matrix tubes prior to homogenization.
* Instruments (e.g., forceps, punch biopsy tool, razor blade) should be sterilized in 100% ethanol between excision of different samples.
* All steps of the procedure should be performed at room temperature. During the procedure, work quickly.

Procedure

1. Excise ~8mg of tissue from a master sample tube, and denote the exact weight.
2. Use forceps to submerge the excised tissue in beads within a lysing matrix tube.
3. Add 350µL of Buffer RLT Plus/ß-Mercaptoethanol/Reagent DX to each lysing matrix tube.
4. Disrupt and homogenize the lysate at 6m/s for 40 seconds.
   1. **NOTE:** Settings stored in the homogenizer as the Human Kidney protocol.
   2. **NOTE**: If excessive foaming occurs in lysing matrix tubes, gently spin down for ~10 seconds prior to proceeding to step 5.
5. Dump the lysate and beads from the lysing matrix tubes into centrifugal filter units.
6. Centrifuge for 3 minutes at 21,100 x g.
7. Carefully transfer supernatant (avoiding pellet) to an AllPrep DNA spin column placed in a 2mL collection tube (supplied).
8. Centrifuge for 30 seconds at 10,000 x g.
   1. **NOTE**: Make sure that no liquid remains on the column membrane following centrifugation. If necessary, repeat the centrifugation until all liquid has passed through the membrane.
9. Place the AllPrep DNA spin column in a new 2mL collection tube (supplied), and store at room temperature for later DNA purification in steps 18-21. Use the flow-through for RNA purification in steps 10-17.
   1. **NOTE**: Do not store the AllPrep DNA spin column at room temperature for long periods. Do not freeze the column.

Total RNA purification

1. Add 350µL of 50% ethanol to the flowthrough from step 9, and mix well by pipetting. Do not centrifuge. Proceed immediately to step 11.
   1. **NOTE**: Do not create bubbles/foaming while mixing with the pipette.
2. Transfer up to 700µL of the sample, including any precipitate that may have formed, to an RNeasy spin column placed in a 2mL collection tube (supplied), and centrifuge for 15 seconds at 12,000 x g. Discard the flowthrough, and reuse the collection tube in step 12.
3. Add 700µL of Buffer RW1 to the RNeasy spin column, and centrifuge for 15 seconds at 12,000 x g. Discard the flowthrough, and reuse the collection tube in step 13.
   1. **NOTE**: After centrifugation, ensure the RNeasy spin column does not contact the flowthrough. Be sure to empty the collection tube completely.
4. Add 500µL of Buffer RPE to the RNeasy spin column, and centrifuge for 15 seconds at 8,000 x g. Discard the flowthrough, and reuse the collection tube in step 14.
5. Add 500uL of Buffer RPE to the RNeasy spin column, and centrifuge for 2 minutes at 8,000 x g. Place the RNeasy spin column in a new 2mL collection tube (supplied).
6. Centrifuge for 1 minute at 21,100 x g.
7. Place the RNeasy spin column in a new 1.5mL collection tube (supplied), and add 50µL of RNase-free water directly to the spin column membrane. Allow the RNase-free water to interact with the spin column membrane for 10 minutes, and, then, centrifuge for 1 minute at 8,000 x g.
   1. **NOTE**: Final eluate can be stored at -70˚C for months.

Genomic DNA purification

1. Add 500µL of Buffer AW1 to the AllPrep DNA spin column from step 9, and centrifuge for 15 seconds at 10,000 x g. Discard the flowthrough, and reuse the spin column in step 18.
2. Add 500µL of Buffer AW2 to the AllPrep DNA spin column, and centrifuge for 2 minutes at 21,100 x g.
   1. **NOTE**: After centrifugation, ensure the AllPrep DNA spin column does not contact the flowthrough. If the column contacts the flowthrough, empty the collection tube, and centrifuge the spin column, again, for 1 minute at 21,100 x g.
3. Place the AllPrep DNA spin column in a new 1.5mL collection tube (supplied), and add 35µL of Buffer EB directly to the spin column membrane. Allow the Buffer EB to interact with the spin column membrane for 1 minute, and, then, centrifuge for 1 minute at 8,000 x g.
4. Repeat step 19 with 20uL of Buffer EB.
   1. **NOTE**: Final eluate can be stored at -70˚C for months.